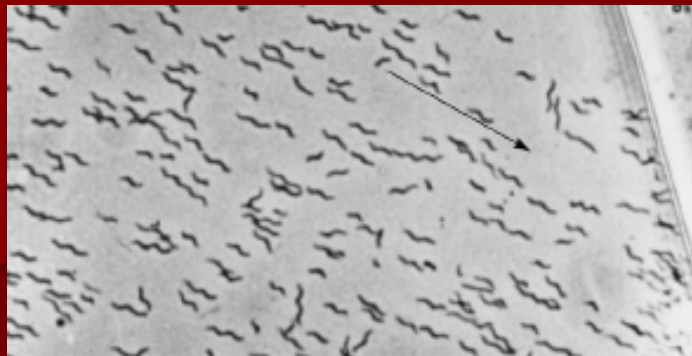


# BACTERIAL MAGNETOSOMES

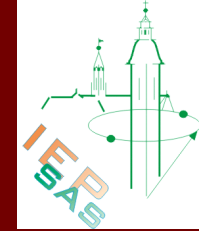
*Magnetospirillum* sp. strain AMB-1



Anežka Džarová

*supervisor:* Dr M. Timko

Our research interests in the section of the magnetism:

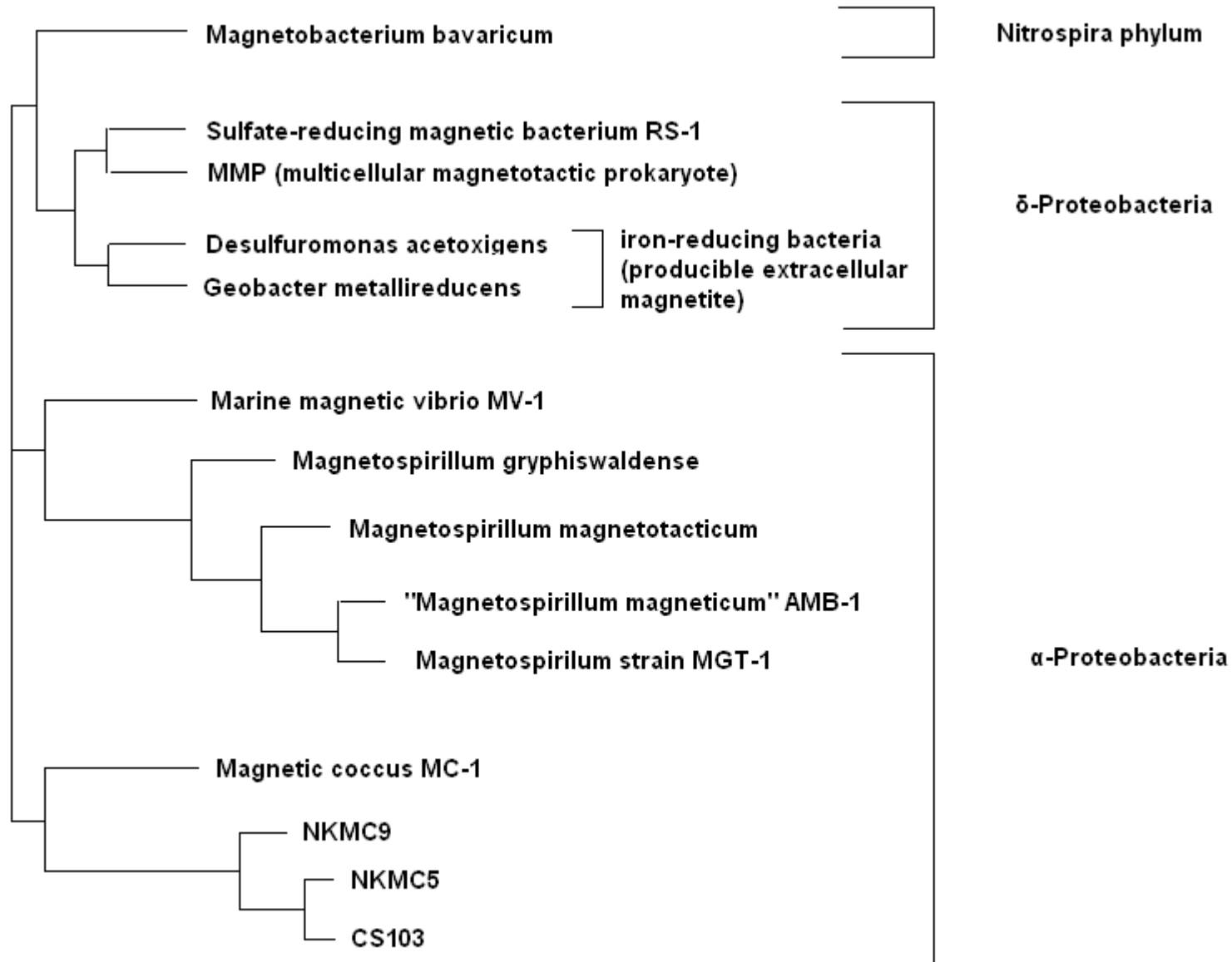


- preparation magnetic particles :

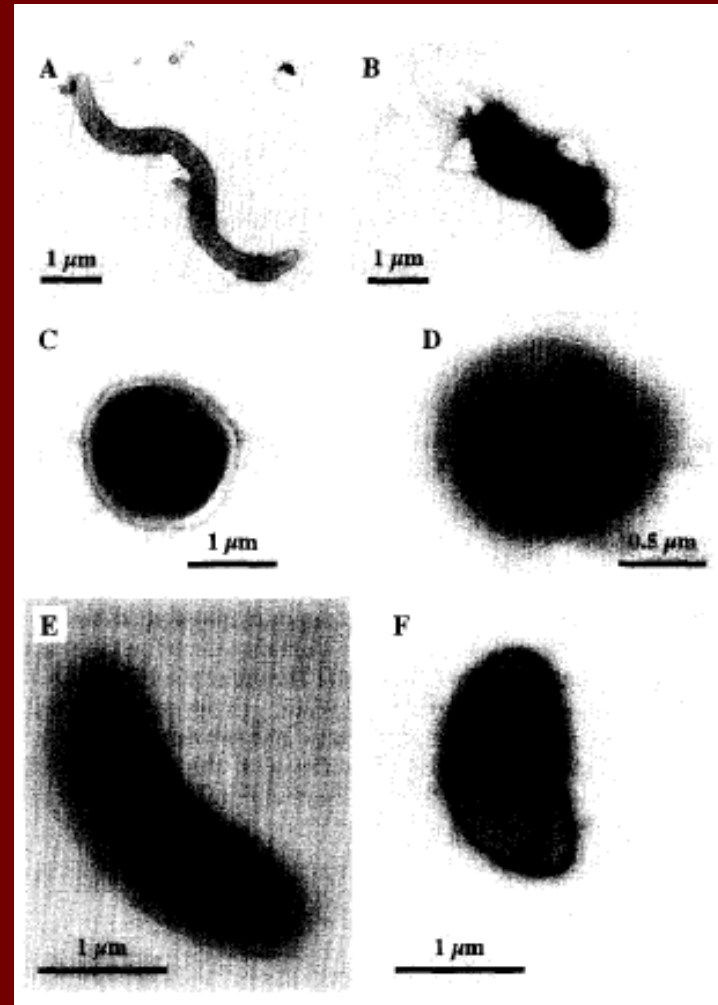
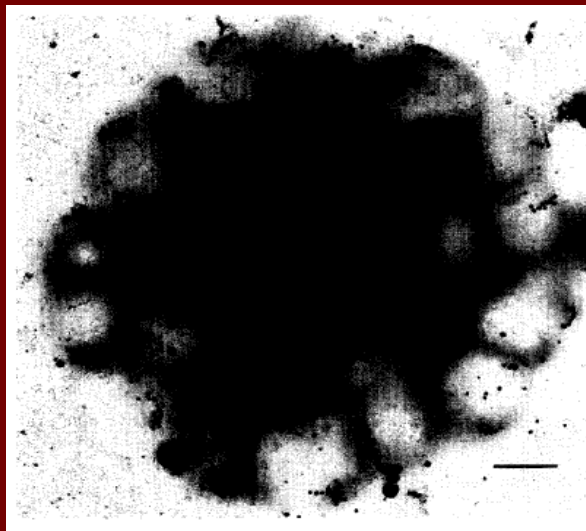
### ***biomineralizations (natural mag. particles)***

- a new biological magnetic particles (magnetosomes) was found as a product of biomineralization process from magnetotactic bacteria. The encapsulation of magnetosomes within the organic membrane provides a natural coating, which ensures superior dispersibility of the  $\text{Fe}_3\text{O}_4$  particles and provides excellent target for immobilization biologically active substances

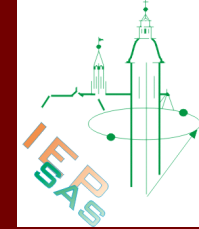
# Dendrogram



- Blakemore (1975)
- they are constituents of natural microbial communities in sediments and chemically stratified water columns and a broad diversity of morphological forms has been found in many marine and freshwater habitats



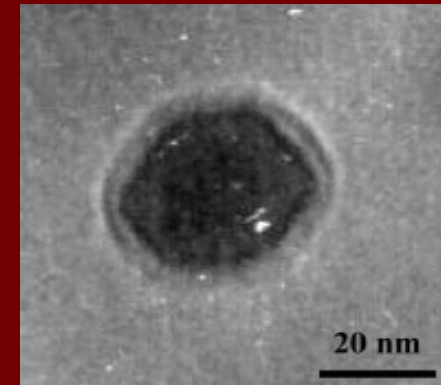




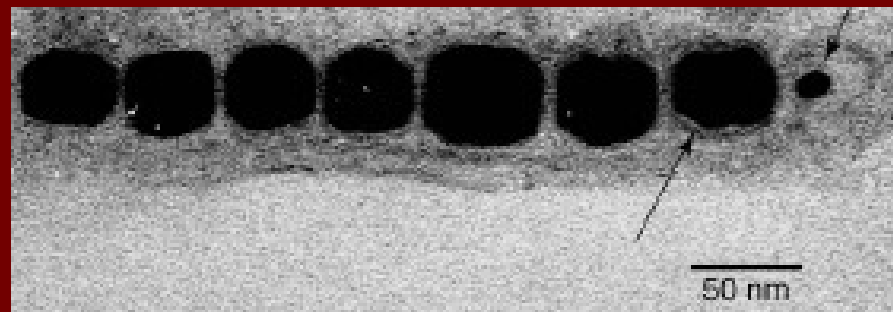
- MTB orient and migrate along geomagnetic field lines – this ability is based on intracellular magnetic structures known as –

## MAGNETOSOMES

[an intracellular single-magnetic-domain crystal of a magnetic iron mineral (the magnetite) to be enclosed by a membrane (a lipid bilayer admixed with proteins) the membrane is intracellular and may be connected to the cytoplasmic membrane]



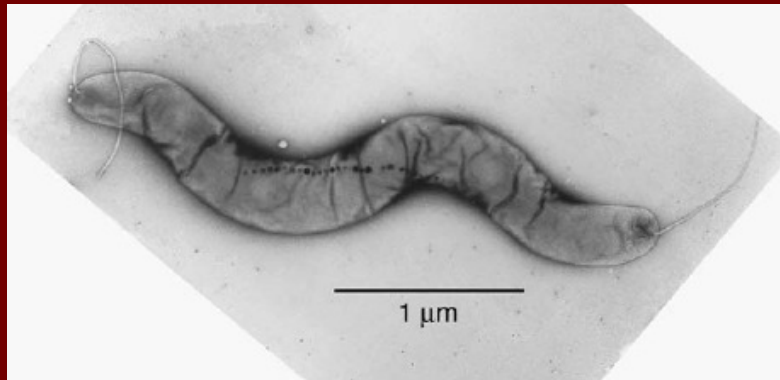
- AMB-1 have single chain of magnetosomes longitudinally traverses the cell



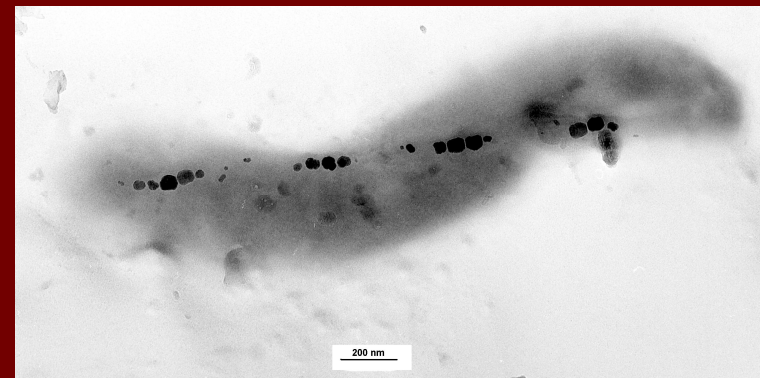
In my experiments magnetosomes were prepared from MTB strain Magnetotacticum Magnetospirillum :

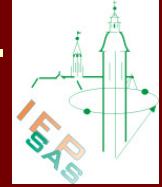


- heterogeneous group of Gram-negative bacteria
- group of aquatic prokaryotes



- strain AMB-1 is Gram-negative  $\alpha$ -proteobacterium that more oxygen-tolerant



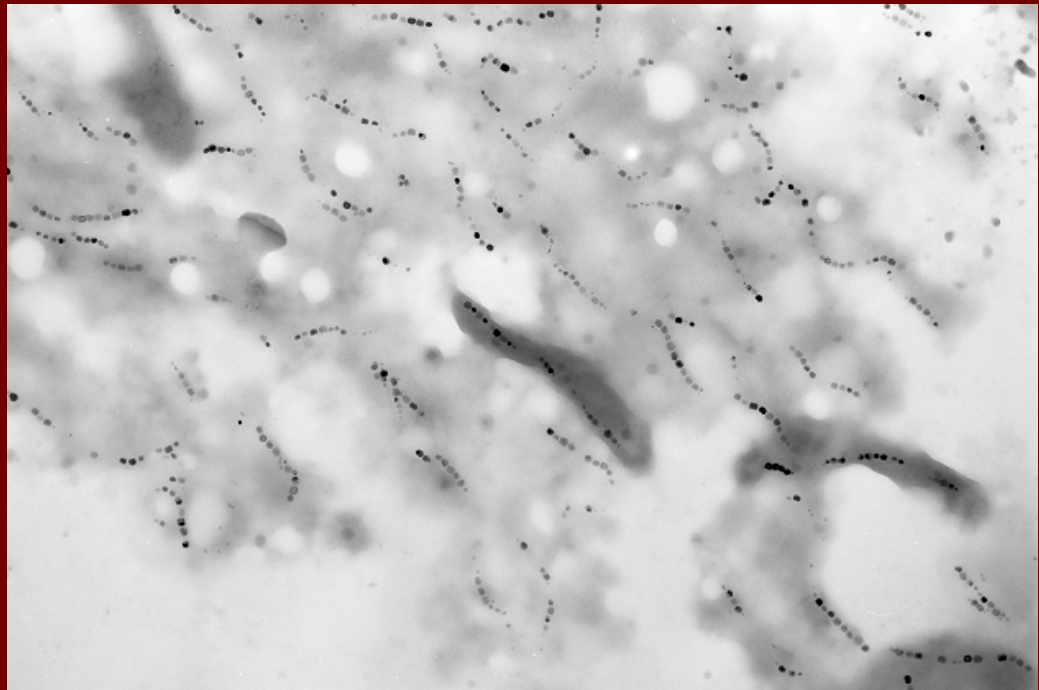
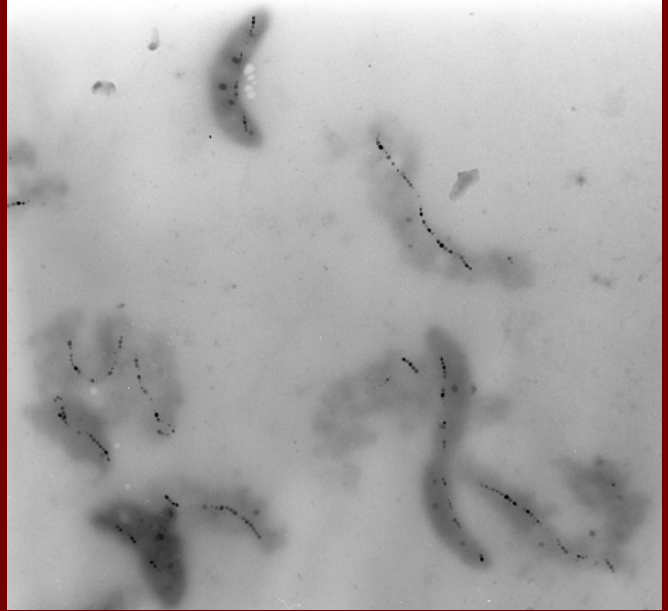
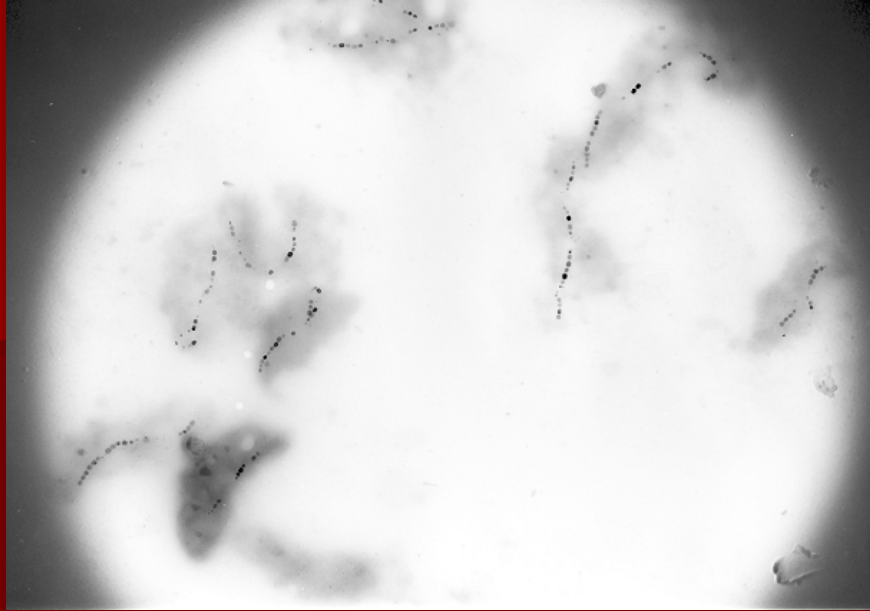


In our experiment for cultivation *Magnetotacticum Magnetospirillum* sp. AMB-1 we used medium consisted of (per 1 L medium):

- 10 mL Wolfe's vitamin solution
- 5 mL Wolfe's mineral solution
- 0.68 g  $\text{KH}_2\text{PO}_4$
- 0.848 g sodium succinate hexahydrate
- 0.575 g sodium tartrate dihydrate
- 0.083 g sodium acetate trihydrate
- 0.225 mL 0.2% (w/v) resazurin (aqueous)
- 0.17 g  $\text{NaNO}_3$
- 0.04 g ascorbic acid
- 2 mL 0.01 M ferric quinate (quinic acid +  $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ )



- resazurin was added to media as colorimetric indicator of redox potential
- the pH was adjusted to 6.75 with NaOH
- this medium was prereduced under nitrogen for a period of 1 hour, using copper as a reducing agent, and was subsequently dispensed into culture tubes in an anaerobic hood
- inoculated tubes were incubated at 25°C for a period of 4 days



## Techniques for the isolation and purification of magnetosome particles from *Magnetotacticum Magnetospirillum* species:

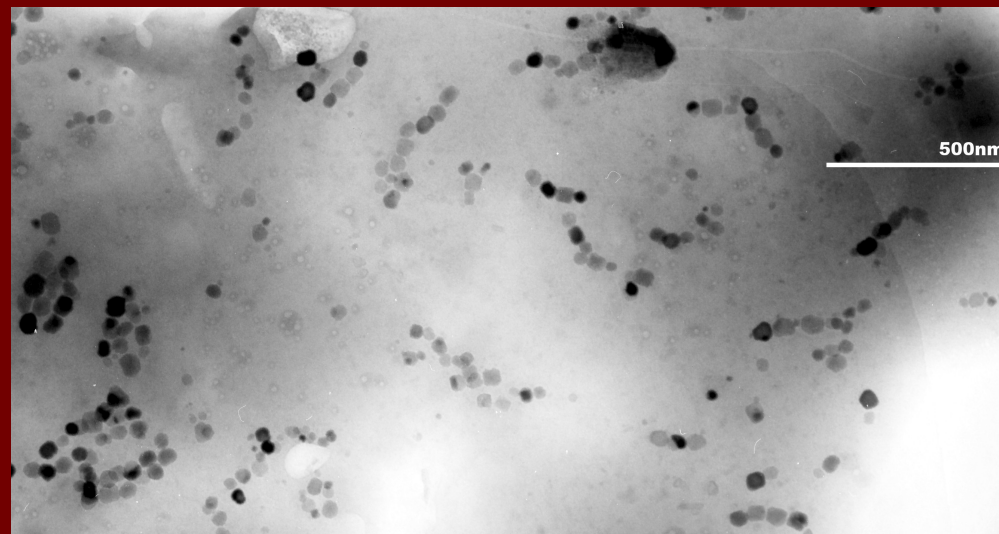
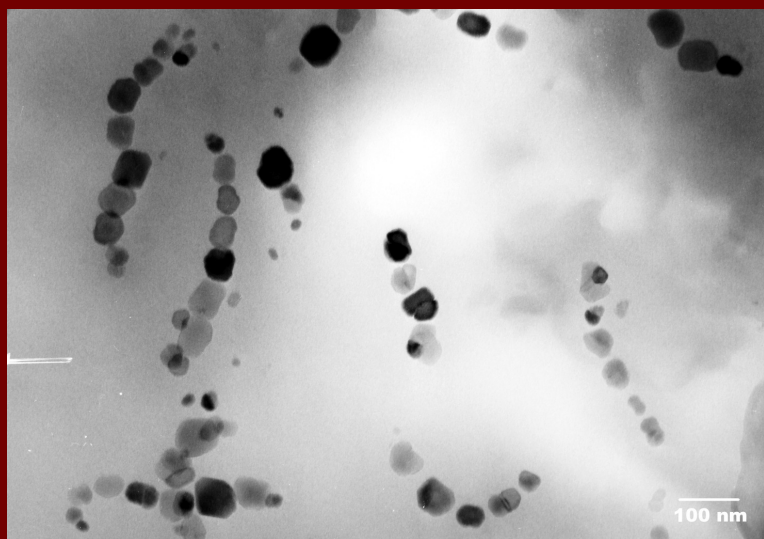
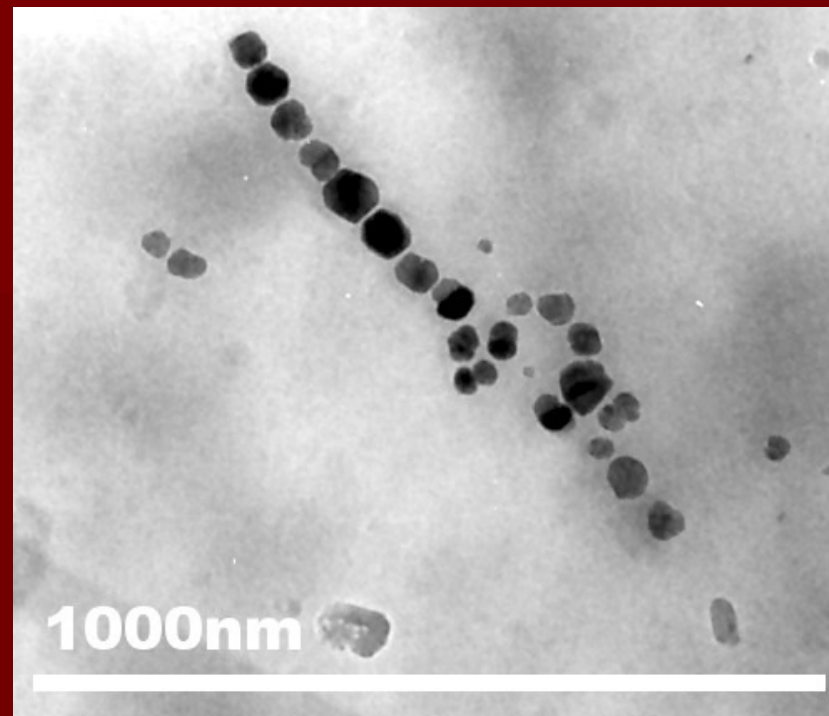
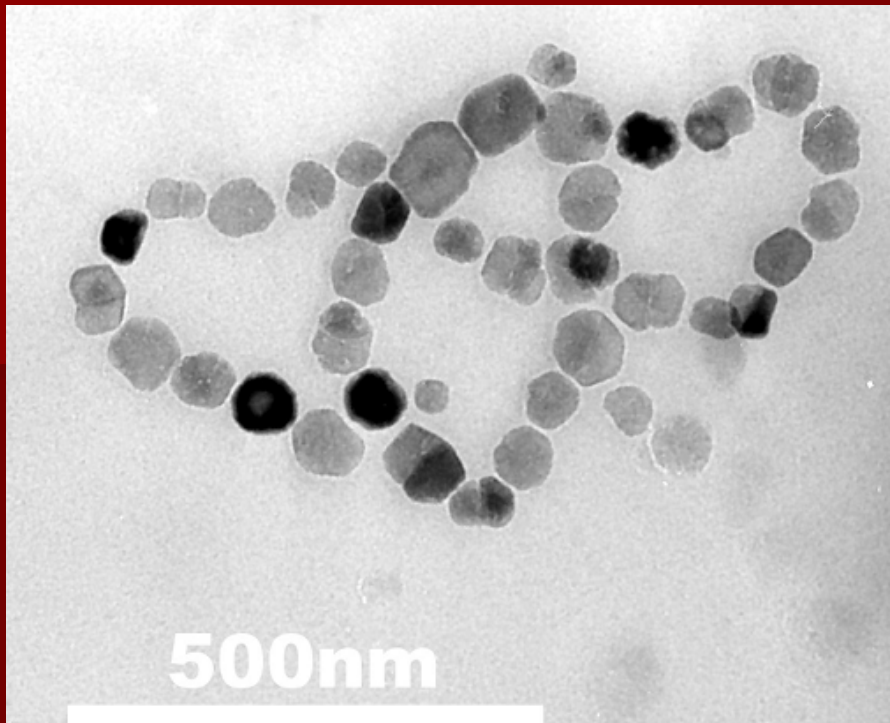


Karen Grünberg, Cathrin Wawer, Bradley M. Tebo, Dirk Schüler, *A large Gene Cluster Encoding Several Magnetosome Proteins Is Conserved in Different Species of Magnetotactic Bacteria*, App. Environ. Microbiol. 2001, 67(10):4573-4582

- are based on magnetic separation or a combination of a sucrose-gradient centrifugation and a magnetic separation technique



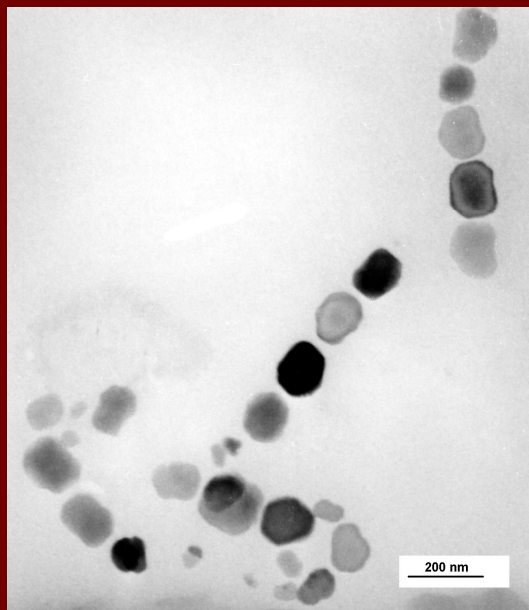




## ***MAGNETOSOMES:***

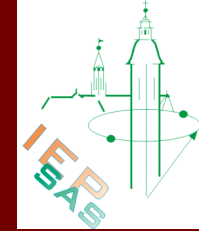


The prepared magnetosomes in our laboratory were examined by TEM, XRD, IČ, SEM, DLS (Dynamic light scattering) and magnetic measurement were examined by SQUID magnetometer Quantum Design:

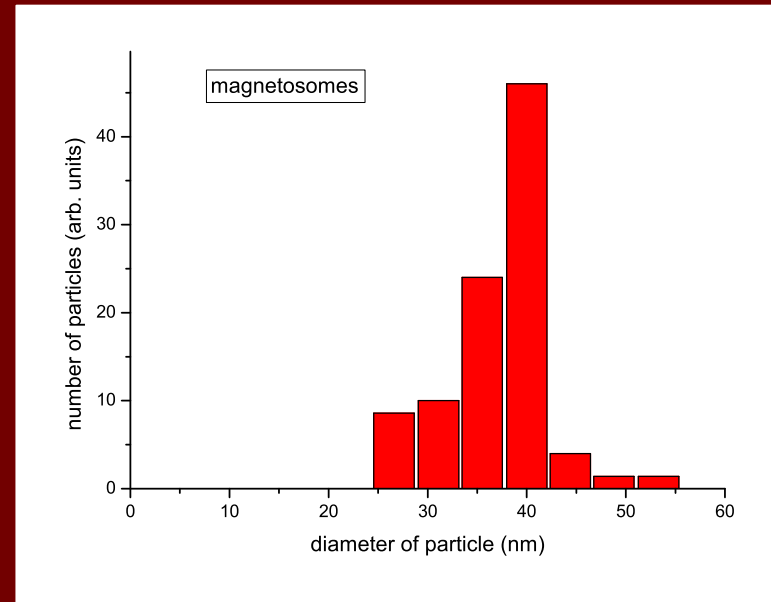
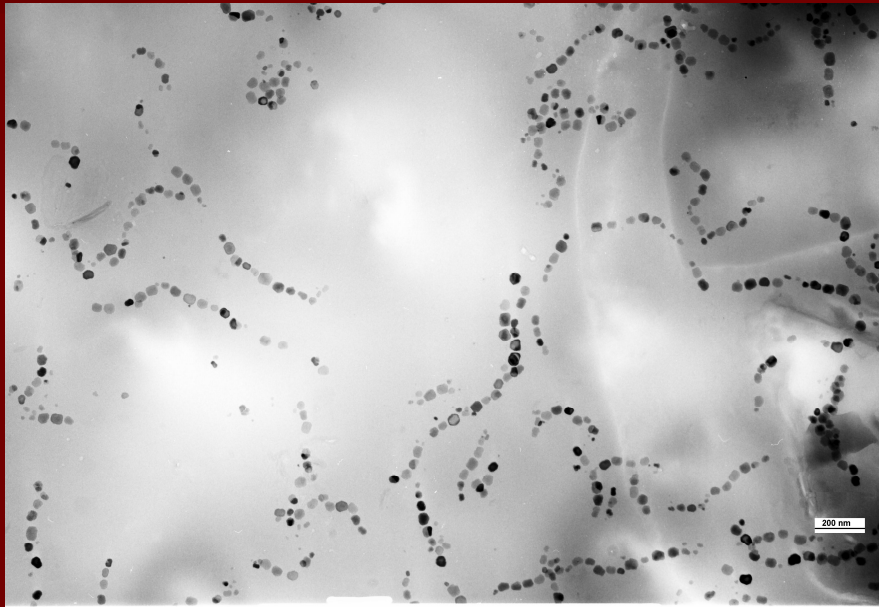


- comprise nanometer –sized, membrane-bound crystals (bacterial magnetic particles) of the magnetic iron minerals magnetite ( $\text{Fe}_3\text{O}_4$ )

# TRANSMISSION ELECTRON MICROSCOPY



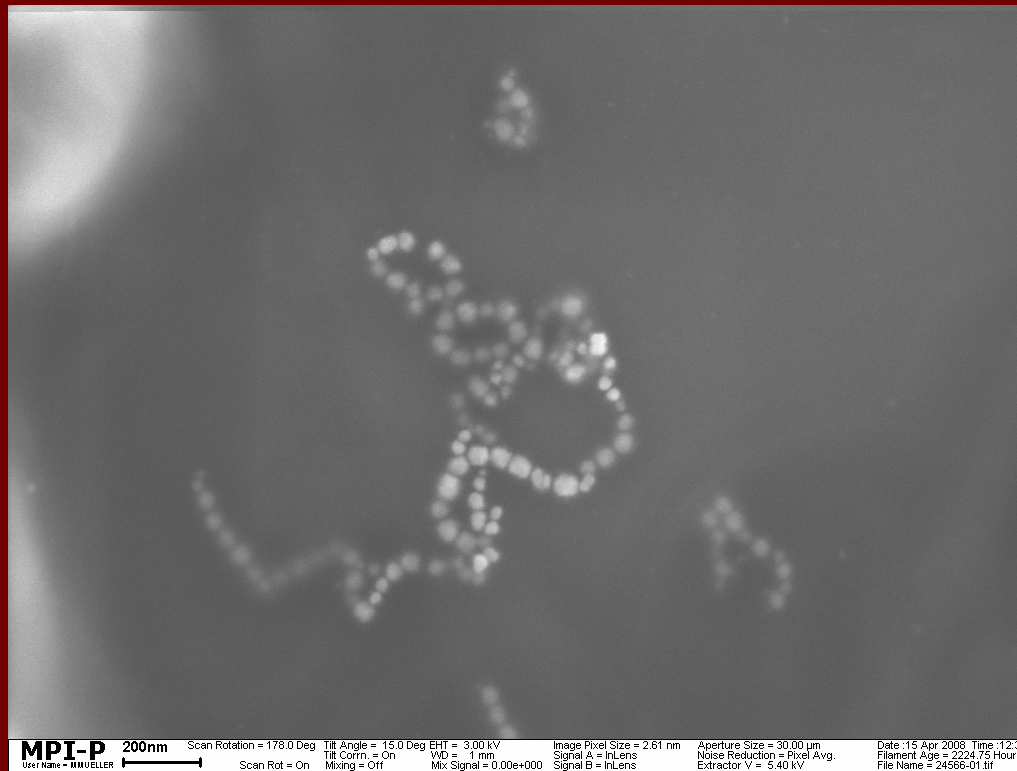
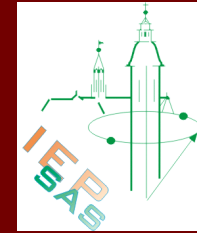
- the mean size of our magnetosomes estimated from TEM was 34 nm





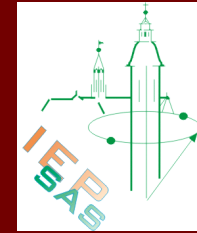
# SCANNING ELECTRON MICROSCOPY

- the mean size of our magnetosomes estimated from SEM was 35 nm



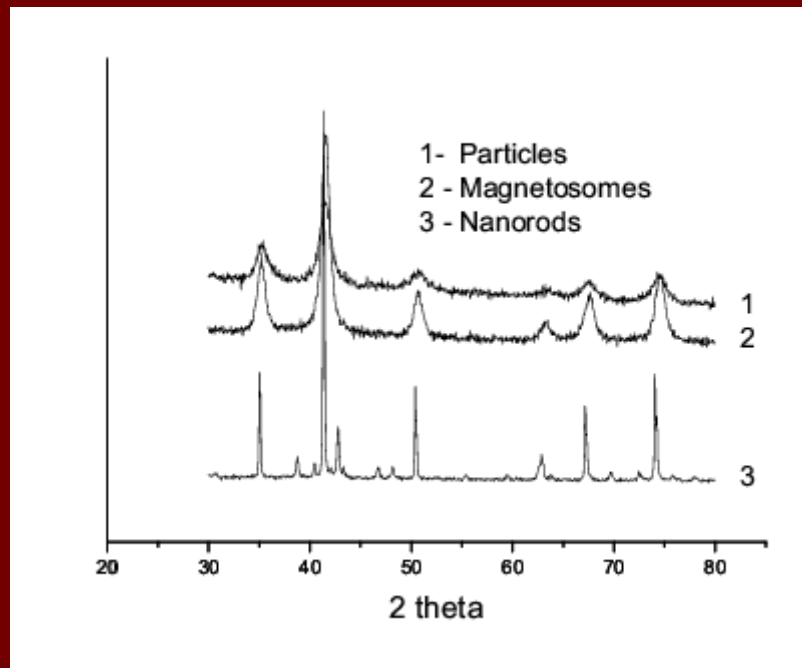
## X-RAY DIFFRACTION

- the mean size of our magnetosomes estimated from and XRD 37 nm



SCHERRER EQUATION:

$$D = 0.9 \lambda / \beta \cos \theta$$

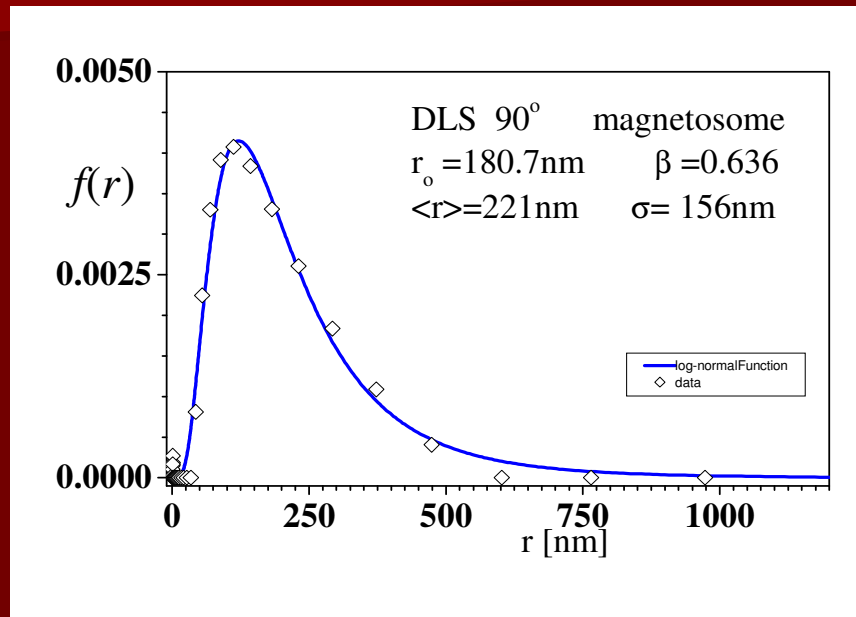
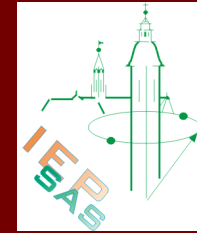


$\lambda$  - wavelength of the incident X-ray

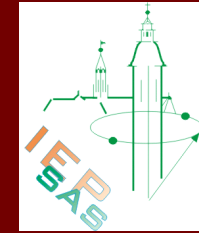
$\theta$  - diffraction angle

$\beta$  - full-width at half-maximum of  
corresponding diffraction peaks

# DLS (Dynamic light scattering)

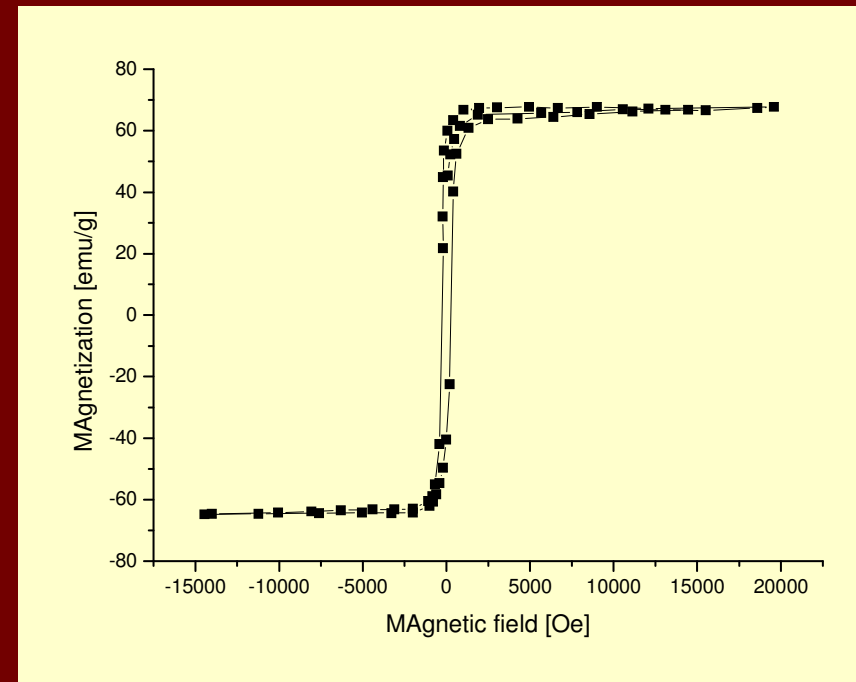


# Magnetic properties were examined by SQUID magnetometer Quantum Design:



- the saturation magnetization of the magnetosomes was estimated to be 62 emu/g what is smaller than for chemically synthesized magnetite 75 emu/g at room temperature due to presence of nonmagnetic organic layer

- the curve of field dependence of magnetization at 293 K exhibited the remanence of 21 emu/g



- coercivity of 185 Oe what is connected with fact that the mean diameter (34 nm) is larger than critical size for transition from superparamagnetic to ferrimagnetic behaviour

Cultivation *Magnetotacticum*  
*Magnetospirillum* sp. AMB-1:



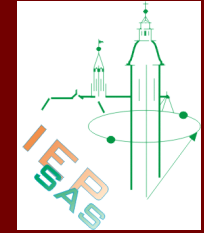






10 mL Wolfe's vitamin solution  
5 mL Wolfe's mineral solution  
0.68 g  $\text{KH}_2\text{PO}_4$   
0.848 g sodium succinate hexahydrate  
0.575 g sodium tartrate dihydrate  
0.083 g sodium acetate trihydrate  
0.225 mL 0.2% (w/v) resazurin (aqueous)  
0.17 g  $\text{NaNO}_3$   
0.04 g ascorbic acid  
2 mL 0.01 M ferric quinate (quinic acid +  $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ )









12.3.09  
2 ml

12.3.09  
2 ml

12.3.09  
2 ml

12.3.09  
2 ml

12.3.09  
2,5 ml

12.3.09  
2,5 ml

12.3.09  
3,5 ml

12.3.09  
9,5 ml

2 ml

2,5 ml

3,5 ml

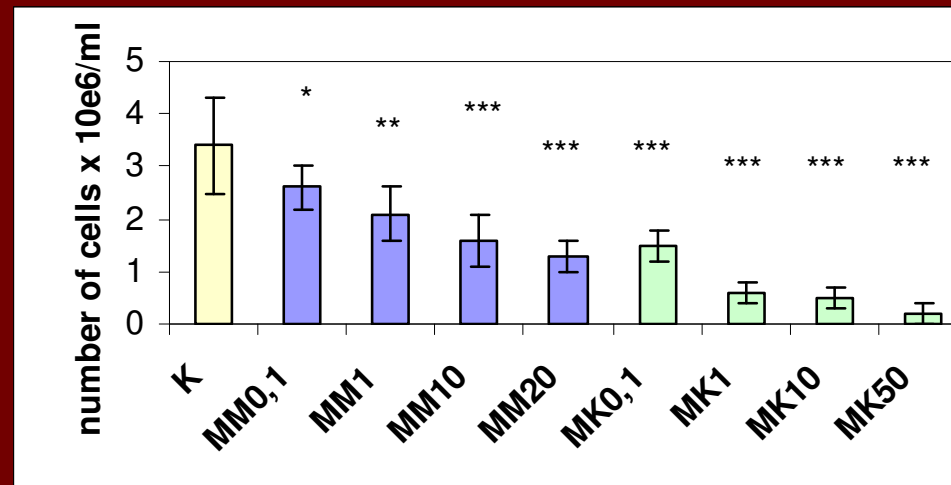
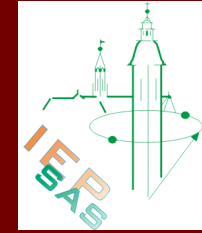
9,5 ml

2 ml

po 6 dňoch

2009 3 18

# The influence of magnetite nanoparticles on human leukocyte activity

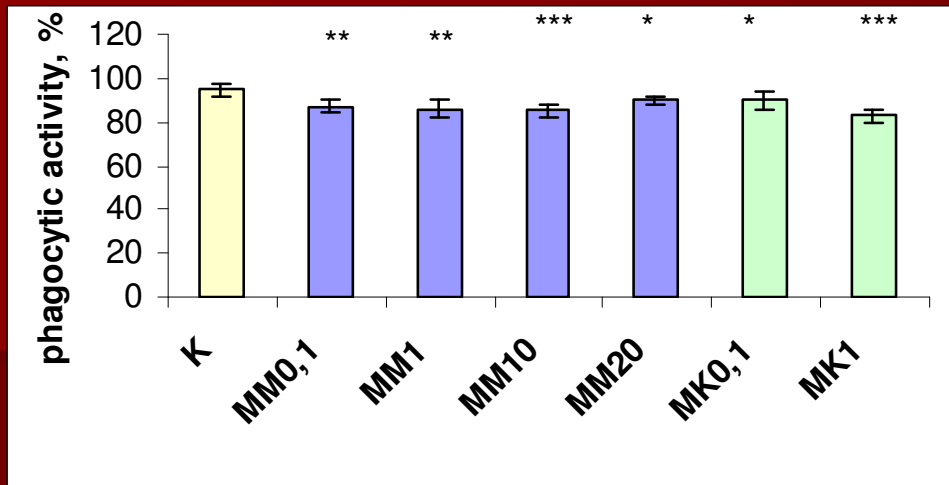


Number of leukocytes/ml for incubation in CO<sub>2</sub> at 37°C

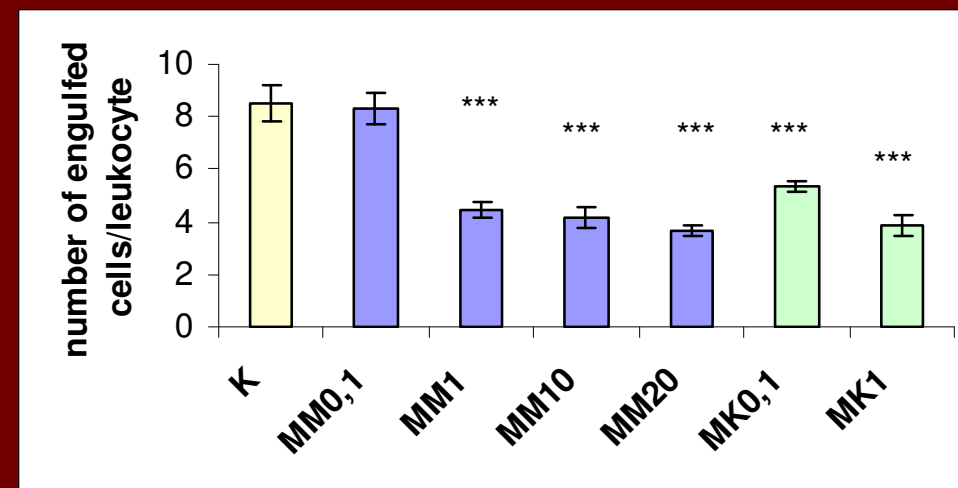
K – control sample

MM – sample of leukocytes with magnetosomes (bacterial magnetite nanoparticles) of concentration: 0,1, 1, 10, 20 µg/ml

MK – sample of leukocyte with synthetic magnetic nanoparticles of concentration: 0,1, 1, 10, 50 µg/ml



Number of phagocytosing cells from 100 phagocytes

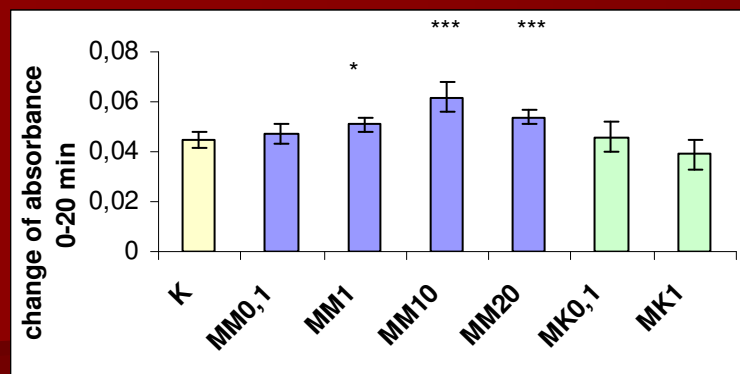
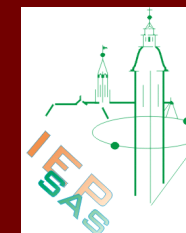


Number of engulfed cells per one leukocyte

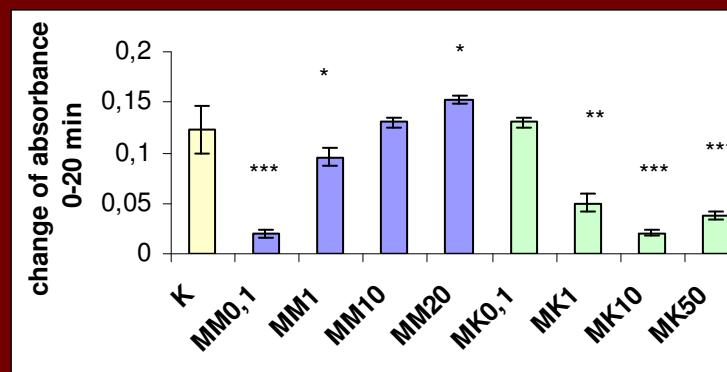
K – control sample

MM – sample of leukocytes with magnetosomes (bacterial magnetite nanoparticles) of concentration: 0,1, 1, 10, 20  $\mu\text{g/ml}$

MK – sample of leukocyte with synthetic magnetic nanoparticles of concentration: 0,1, 1, 10, 50  $\mu\text{g/ml}$



Lysozyme activity expressed change of absorbance (410 nm) after 20 minutes of reaction

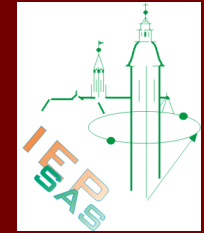


Peroxidase activity expressed change of absorbance (490 nm) after 20 minutes of reaction

K – control sample

MM – sample of leukocytes with magnetosomes (bacterial magnetite nanoparticles) of concentration: 0,1, 1, 10, 20  $\mu\text{g/ml}$

MK – sample of leukocyte with synthetic magnetic nanoparticles of concentration: 0,1, 1, 10, 50  $\mu\text{g/ml}$



## CONCLUSION:

- successful cultivation and isolation
- magnetosome in MTB are arranged in straight chains
- after isolation from MTB those chains tend to form closed loops
- the mean size is 34 nm (TEM, SEM) or 37 nm (XRD)
- saturation magnetization of the magnetosomes  $M_s=62$  emu/g
- coercivity  $H_c=185$  Oe
- remanence  $M_r= 21$  emu/g
- the both tested samples lysed (killed) leukocyte cells during incubation. From obtained results follow that spherical nanoparticles are more aggressive material than magnetosomes

## PUBLICATIONS:

- TIMKO, Milan - DŽAROVÁ, Anežka - ZÁVIŠOVÁ, Vlasta – KOVÁČ, Jozef – ŠPRINCOVÁ, Adriana - KONERACKÁ, Martina - KOPČANSKÝ, Peter – TOMAŠOVIČOVÁ, Natália - GOJZSEWSKI, Hubert – SKUMIEL, Andrzej – JOZEF CZAK, Arkadiusz – VÁVRA, Ivo Magnetic Properties and Heating Effect in Bacterial Magnetic Nanoparticles. In. ***Journal of Magnetism and Magnetic Materials***, 321 (2009) 1521–1524
- KOPČANSKÝ, Peter - TOMAŠOVIČOVÁ, Natália - KONERACKÁ, Martina - ZÁVIŠOVÁ, Vlasta - TIMKO, Milan - DŽAROVÁ, Anežka - ŠPRINCOVÁ, Adriana - ÉBER, N. - FODOR-CSORBA, K. - TÓTH-KATONA, T. - VAJDA, A. - JADZYN, Jan. Structural changes in the 6CHBT liquid crystal doped with spherical, rodlike, and chainlike magnetic particles. In ***Physical Review E***. ISSN 1539-3755, 2008, vol. 78, part 1, p. 011702-1-5.
- TIMKO, Milan - DŽAROVÁ, Anežka - KOPČANSKÝ, Peter - ZÁVIŠOVÁ, Vlasta - KONERACKÁ, Martina - KOVÁČ, Jozef - ŠPRINCOVÁ, Adriana - VACLAVÍKOVÁ, Miroslava - IVANIČOVÁ, Lucia - VÁVRA, Ivo. Magnetic Properties of Magnetite Formed by Biomineralization and Chemical Synthesis. In ***Acta Physica Polonica A***. ISSN 0587-4246, 2008, vol. 113, no. 1, p. 573-576.



- TIMKO, Milan - DŽAROVÁ, Anežka - KOVÁČ, Jozef – KOPČANSKÝ, Peter - GOJZSEWSKI,, Hubert – SZLAFEREK , Andrzej. Magnetic Properties of Bacterial Nanoparticles, In ***Acta Physica Polonica A***, 2009, vol 115, no. 1, 381-383

- TIMKO, Milan - DŽAROVÁ, Anežka - ZÁVIŠOVÁ, Vlasta - KONERACKÁ, Martina - ŠPRINCOVÁ, Adriana - KOPČANSKÝ, Peter - KOVÁČ, Jozef - VÁVRA, Ivo - SZLAFEREK, A. Magnetic properties of bacterial magnetosomes and chemosynthesized magnetite nanoparticles. In ***Magnetohydrodynamics***. ISSN 0024-998X, 2008, vol. 44, no. 2, p. 113-120.

- DŽAROVÁ, Anežka - TIMKO, Milan - ŠPRINCOVÁ, Adriana - KOPČANSKÝ, Peter - KOVÁČ, Jozef - KONERACKÁ, Martina - VACLAVÍKOVÁ, Miroslava - VÁVRA, Ivo. Formation and magnetic properties of magnetosomes. In ***Materials Structure in Chemistry, Biology, Physics and Technology***. ISSN 1211-5894, 2008, vol. 15, no. 1, p. 10-12.





**Thank you for your attention**

